

Review Article:**MicroRNAs in health and disease****K. Rama, P.V.L.N. Srinivasa Rao, Aparna R. Bitla***Department of Biochemistry, Sri Venkateswara Institute of Medical Sciences, Tirupati***ABSTRACT**

MicroRNAs (miRNAs) are a class of endogenous small non-coding RNAs about 21-25 nucleotides in length and account for 1%-5% of the genome. Ever since its discovery in 1993, these molecules have attracted researchers due to their crucial role in regulating gene expression. Dysregulation in miRNA expression profile could serve as molecular signatures for identifying diseases. These have found applications in a number of diseases including cancer, wherein they may be useful for as biomarkers in cancer diagnosis as well as serve as therapeutic targets. This review provides an update on the evolution of miRNAs in the field of medicine and its future therapeutic applications.

Key words: *MicroRNAs, Biomarkers, Transcriptome*

Rama K, Srinivasa Rao PVLN, Bitla AR. MicroRNAs in health and disease. J Clin Sci Res 2017;6:25-34. DOI: <http://dx.doi.org/10.15380/2277-5706.JCSR.16.11.006>.

INTRODUCTION

MicroRNAs were discovered during RNA extraction procedures. Ribonucleic acid (RNA) when extracted from tissues or cells showed a pool of small RNA molecules which were assumed to be products of RNA degradation, arising as a result of RNA extraction procedure. These small RNA molecules were later shown to be non-coding RNA molecules involved in gene expression.¹ The small non-coding RNA molecules include small nuclear RNAs (snRNAs) which are involved in mRNA splicing and the small nucleolar RNAs (snoRNAs) which are involved in ribosomal RNA processing. Other classes of non-coding RNA are involved in the silencing of gene expression. These are subdivided into three types: short interfering RNAs (siRNAs) which target mRNA structure, others which target chromatin for epigenetic modification and the micro RNAs (miRNAs) which regulate mRNA translation.¹

MicroRNAs are small non-coding RNAs about 21-25 nucleotides in length accounting for 1% -5% of the genome in plants and animals.² They regulate the expression of genes involved in cell proliferation and apoptosis, development, differentiation, metabolism, immunity, stress response, aging and cell cycle control. Approximately 50% of protein coding genes are regulated by miRNAs in a mammalian genome³ and are located in either the introns of protein coding gene or non-coding region of genes or the intragenic regions of the genome.⁴

Discovery of microRNAs

The first miRNA to be discovered was lin-4 identified during screening for defects in the temporal control of post-embryonic development in *Caenorhabditis elegans* in 1993.⁵ *C.elegans* cell lineages have four different larval stages (L₁-L₄). Lin-4 activity is required for the transition from L₁ to L₂.⁵ Null mutation in Lin-4 causes a failure in temporal

Received: November 29, 2016; Accepted: December 29, 2016.

Corresponding author: Dr P.V.L.N. Srinivasa Rao, Sr. Professor and Head, Department of Biochemistry, Sri Venkateswara Institute of Medical Sciences, Tirupati, India.
e-mail: seenupvln@yahoo.com

**Online access**

http://svimstpt.ap.nic.in/jcsr/jan-mar17_files/ra.16.11.006.pdf
DOI: <http://dx.doi.org/10.15380/2277-5706.JCSR.16.11.006>

development.^{5,6} Animals with this mutation have certain adult structures missing and developmental delays. In 1987, Ferguson et al,⁷ identified a suppressor mutation in the gene *Lin-14* which could revert the null mutations in *Lin-4*. These null mutations led to a phenotype which was opposite to that caused by *Lin-4* null mutations, indicating a negative regulation of *Lin-14* by *Lin-4*.⁸

The second miRNA *let-7* was also discovered in *C.elegans* in the year 2000. *Let-7* encoded a 21 nucleotide RNA which controls the L_4 to adult transition of larval development.⁹ Loss of *let-7* activity causes reappearance of larval cell fates during adult stage of development. *Let-7* activity has been detected in vertebrates, ascidian, hemichordate, mollusk, annelid and arthropods. RNAs from plant and unicellular organisms did not show *Let-7*.¹⁰ *Let-7* expression has also been shown in human tissues including brain, kidney, heart, liver, lung, small intestine, stomach and thymus.¹¹

Nomenclature of microRNAs

MicroRNAs are named using the prefix “miR” and a unique identification number, e.g., miR-1, miR-12 etc. Mature miRNAs differing only in one or two positions are given suffixes; example, miR-10a and miR-10b. The mature sequences are designed as “miR” in the database, whereas the precursor hairpins are labeled as “mir”.¹²

Biogenesis of microRNAs

The biogenesis of miRNA (Figure 1) involves three sequential steps:^{1,13} (i) transcription: RNA polymerase II is involved in transcription of majority of miRNAs; (ii) processing of primary miRNA (pri-miRNA) into a precursor miRNA in the nucleus (pre-miRNA); and (iii) Maturation of pre-miRNA into a functional miRNA in the cytoplasm.

The primary miRNA transcripts (pri-miRNAs) are several hundred nucleotides long and characterized by the formation of stem loop

structures. Modification of pri-miRNAs occurs similar to that of protein-coding transcripts i.e., addition of a 5' cap and a 3' poly-A-tail. The enzyme “drosha” then processes the pri-miRNAs into pre-miRNAs in the nucleus. Pre-miRNA is usually 60-100 nucleotides long and is exported from the nucleus in a RAS-related nuclear protein (Ran) Guanosine triphosphate (GTP)-dependent process after binding with exportin-5.¹⁴ The pre-miRNA is further processed in the cytoplasm, by the enzyme dicer into a 19-24 nucleotide long double stranded mature miRNA. The two strands of this miRNA are named the ‘guide strand’ and the ‘passenger strand’. The guide strand incorporates into the RNA induced silencing complex (RISC) while the passenger strand is unwound from the guide strand and undergoes degradation¹³ or both strands may remain functional.¹⁵ The complex formed after incorporating miRNA guide strand with RISC is known as a ‘miRISC complex’.¹ This complex binds to the 3' untranslated region of target mRNA and induces mRNA degradation or represses

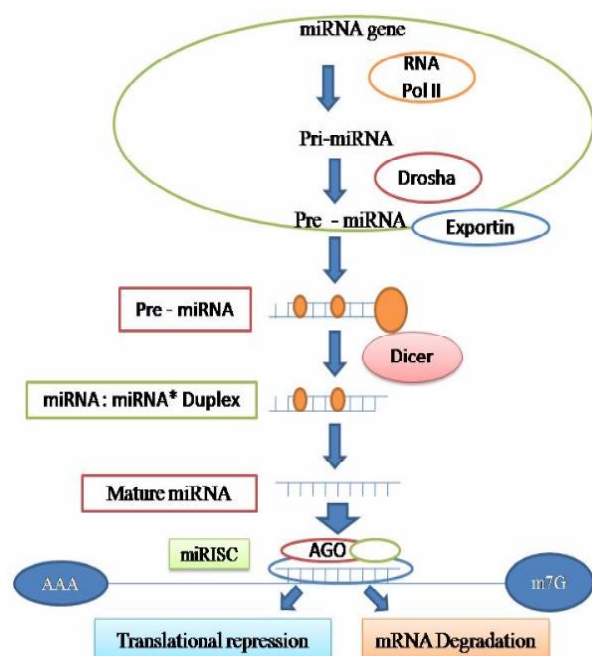


Figure 1: Biogenesis of miRNAs

miRNA=microRNA, RNA=Ribonucleu acid; miRISC=micro RNA-induced silencing complex; AGO=arogonuate protein; AAA=polyA tail; m7G=7-methyl guanosine

translation (Figure 1) depending on the degree of sequence complementarity between miRNA and mRNA target.¹⁶

Circulating miRNAs

Most of the microRNAs are found intracellularly. However, a number of miRNAs have been observed in extracellular locations including their presence in various body fluids such as serum, plasma, tears, breast milk, bronchial lavage, colostrum, seminal, amniotic, pleural, peritoneal and cerebrospinal fluids.^{3,17-19} These miRNAs are more stable than RNAs, since they are shielded from ribonuclease degradation by packaging in lipid vesicles such as microvesicles, exosomes, in combination with RNA-binding proteins or both.^{20,21} MiRNAs have been identified in cell derived lipid vesicles, microvesicles which are relatively large (100 nm to 1 μ) and exosomes which are smaller (30-100 nm).¹³ These extracellular miRNAs are involved in cell-cell communications.^{22,23}

Circulating miRNAs are highly stable even when subjected to extremes of pH, temperature, extended storage and multiple freeze-thaw

cycles.³ They have been isolated successfully from clinical specimens such as sputum,²⁴ plasma^{25,26} and serum.²⁷ This makes them suitable to be used as non-invasive biomarkers.^{26,28}

The expression profiles of circulating miRNAs have been studied in non-malignant and malignant diseases to identify disease-specific expression patterns or miRNA signatures.^{13,28} These miRNA signatures can be used to discriminate healthy controls from patient samples with a high level of accuracy. Thus, circulating miRNAs in blood specimens could serve as diagnostic markers.²⁹

Circulating microRNAs in health and disease

Dysregulation of specific miRNAs in the form of either up-regulation or down-regulation have been reported in a number of diseases including malignancies, cardiovascular diseases, skin diseases and autoimmune diseases (Table 1).

MicroRNAs in cardiovascular diseases

MicroRNAs have been shown to play a diagnostic and therapeutic role in

Table 1: Expression of miRNAs in human diseases

Disease	Up-regulated	Down-regulated
Cardiovascular Disease		
Coronary artery disease	miR-1 ³⁴	
Cardiac hypertrophy		miR-29 ³⁴ , miR-133 ³⁴
Atherogenesis		miR - 126 ³⁴
Alzheimer	miR-9, ⁵⁰ miR-128a, ⁵⁰ miR-125b ⁵⁰	
Psoriasis	miR-203, ⁵⁵ miR - 146 ⁵⁵	let-7d, ⁵⁵ miR-142-3p and miR-181a
Dermatomyositis		miR - 223 ⁵⁹
Rheumatoid arthritis	miR-155, ⁶⁷ miR-146 ⁶⁷	
Systemic lupus erythematosus	miR-189, ⁶⁸ miR-61, ⁶⁸ miR-78, ⁶⁸ miR-21, ⁶⁸ miR-142-3p, ⁶⁸ miR 342, ⁶⁸ miR-299-3p, ⁶⁸ miR-198 ⁶⁸ and miR-298 ⁶⁸	miR-196a, ⁶⁸ miR-17-5p, ⁶⁸ m 409-3p, ⁶⁸ miR-141, ⁶⁸ miR-3 miR - 112, ⁶⁸ and miR-184 ⁶⁸
Breast cancer	miR-125b, ⁷⁷ miR-145, ⁷⁷ miR-21, ⁷⁷ and miR-155 ⁷⁷	
Colorectal cancer		miR-143 ⁷⁸ and miR-145 ⁷⁸
Papillary thyroid cancer	miR - 21 ⁸⁵	

miR=micro RNA; Let-7d=Lethal-7d

cardiovascular diseases. A number of miRNAs including miR-1, miR-16, miR-27b, miR-30d, miR-126, miR-133, miR-143, miR-208 and the let-7 family are expressed in healthy cardiac tissue and play a role in normal cardiac maintenance as well as in diseases of the heart.³⁰ The miR-143/-145 (miR-143/-145) which are abundantly expressed in vascular smooth muscle cells (VSMCs) have been shown to be involved in the differentiation of VSMCs and determine the VSMC phenotypic switching.³¹⁻³³ They could thus serve as potential drug targets for vascular diseases such as atherosclerosis, hypertension, and CAD.³³

Other miRNAs implicated in causing cardiovascular diseases include miR-1 in arrhythmia, miR-29 in cardiac fibrosis, miR-126 in angiogenesis and miR-133 in cardiac hypertrophy.³⁴ The miR-1 has been shown to have a role in cardiac morphogenesis, controlling cell fate of different lineages and thus helping in normal development of cardiac chambers.³⁵ miR-1 has been reported to be over expressed in individuals with coronary artery disease,³⁶ It thus represents a potential drug target in patients with arrhythmias.

The role of miR-126 in regulating endothelial cell biology and maintaining vascular structure has been demonstrated.³⁷ It was shown to target sprouty-related Extractable Nuclear Antigen/ Vasodilator-stimulated phosphoprotein (VASP) Homology protein 1 (EVH1) domain-containing protein 1 (*SPRED1*) a protein that negatively downregulates mitogen activated protein (MAP) kinase pathway, vascular cell adhesion molecule 1 (*VCAM1*) and phosphoinositol-3 kinase regulatory subunit 2 (*PIK3R2*) for repression and functions to promote vascular endothelial growth factor (VEGF) signaling by inhibiting *SPRED1* and *PIK3R2*.³⁷ A study demonstrated the beneficial effects of aerobic exercise training on cardiac remodeling through its effect on certain miRNAs.³⁸ The effects included decreasing cardiac fibrosis by

inhibiting collagen through upregulation of miR-29, inhibition of negative regulators of VEGF pathway. These effects of miR-126 in turn increase angiogenesis and modulation of the renin-angiotensin system by miRNAs-27a/b and -143.³⁸

MicroRNAs in neurodegenerative diseases

The miRNAs have been shown to be expressed in the central nervous system and play a role in brain development with specific temporal and spatial patterns of expression.³⁹ A number of microRNAs including let-7B, miR-9, 134,127,137 and 184 have been shown to have a role in neural stem cell proliferation and differentiation.⁴⁰⁻⁴² The miR-79, an epidermal microRNA regulates neuronal migration through control of the cellular glycosylation state,⁴³ while miR-132⁴⁴ mediates the integration of newborn neurons into the adult dentate gyrus. MicroRNAs have been shown to play a fundamental role in migration and integration; processes that are critical for the architecture of the brain.⁴⁵ miRNAs especially miR-9,⁴⁶ miR-132⁴⁷ and miR-134⁴⁸ have been shown to play a regulatory role in dendritic development and maturation in experimental models.

Neurodegenerative diseases related to aging are the result of different genetic and environmental influences.⁴⁹ Dysregulation in miRNA expression has been reported in Alzheimer's, Parkinson's and Huntington's disease. Dysregulation of miR-9, miR-125b, miR-128a, and miR-124a has been reported in Alzheimer brain samples when compared with age-matched controls among a subset of 12 miRNAs studied.⁵⁰ The miR-9 was found to be up-regulated in both fetal hippocampus and AD hippocampus, miR-128a in AD while miR-125b showed an increasing non-significant trend in AD.⁵⁰ The authors⁵¹ studied the same set of miRNAs in cultured human fetal brain-derived primary neural (HN) cells, including astrocytes and neurons, treated with metal salts,

such as aluminum and iron sulfates to stimulate reactive oxygen species (ROS). ROS increased the expression of miR-9, miR-128 and, to a lesser extent, miR-125b. HN treated cell compared to controls supporting the hypothesis that ROS influence AD brain through pathways specifically mediated by miRNAs.⁵¹

Using Purkinji cells, the role of miRNAs in the survival of differentiated neurons through conditional Purkinje cell-specific ablation of Dicer, the key enzyme involved in miRNA generation causing Purkinje cell death has been demonstrated.⁵² Apart from their role in neurodevelopment, miRNAs have been shown to be useful biomarkers.^{53,54} The role of miRNAs in development of neurodegenerative diseases as well as use as biomarkers make them potential targets for treating neurodegenerative diseases.

MicroRNAs in skin disease

Dysregulation in miRNA expression has also been shown in skin diseases. An up-regulation of miR-203 which is expressed exclusively in keratinocytes in patients with psoriasis-affected skin compared to healthy skin has been reported.⁵⁵ The miR-146 was also found to be upregulated in psoriasis patients.⁵⁵ The miR-146 has been shown to be a NF-kappaB-dependent gene involved in cytokine signalling. It has been shown to down-regulate IL-1 receptor-associated kinase and tumour necrosis factor (TNF) receptor-associated factor 6 protein levels through a negative feedback regulation loop.⁵⁶ Circulating miRNAs dysregulated in psoriasis include miR-128a (up-regulated), let-7d, miR-142-3p and miR-181a (down-regulated). Treatments using anti-TNF agent etanercept showed decrease in miR-106b, miR-26b, miR-142-3p, miR-223 and miR-126 in responders.⁵⁷

Circulating miRNA-221 has been proposed as a tumor marker in patients with malignant melanoma.⁵⁸ It has been shown to differentiate in-situ malignancy from stage I-IV and

correlated with tumor thickness.⁵⁸ Serum miRNA-223 levels have been reported to be lower in patients with dermatomyositis.⁵⁹

MicroRNAs in autoimmune diseases

MicroRNAs have a role in regulating immune response and prevent autoimmunity. As discussed above miRNA-146 is involved in down-regulating inflammatory response.⁵⁵ Other miRNA involved in regulating the innate immunity to bacterial and viral pathogens is miRNA-155.⁶⁰ This action is through its binding to Src homology-2 domain-containing inositol 5-phosphatase 1 (SHIP1) and suppressor of cytokine signaling 1 (SOCS1) targets which are negative regulators of the immune response.⁶⁰ MiRNAs are also involved in regulating adaptive immune response^{61,62} through the development and differentiation of T and B lymphocytes.^{63,64}

Studies have reported an association of miRNAs with autoimmune inflammatory conditions like rheumatoid arthritis (RA). Increased expression of miR-155 and miR-146 have been reported in synovial fibroblasts and synovial tissue^{65,66} and peripheral blood mononuclear cells in RA.⁶⁷ Dysregulation in miRNA expression has also been reported in SLE, another autoimmune disease characterized by overproduction of auto antibodies towards several self antigens. Using microarray assay for miRNA expression in polymorphonuclear leukocytes, down regulation of seven miRNAs including miR-196a, miR-17-5p, miR-409-3p, miR-141, miR-383, miR-112, and miR-184 and up-regulation of nine miRNAs namely miR-189, miR-61, miR-78, miR-21, miR-142-3p, miR-342, miR-299-3p, miR-198, and miR-298 has been reported.⁶⁸ Similarly, dysregulation of 66 miRNAs was described in patients with lupus nephritis.⁶⁹

MicroRNAs in cancer

MiRNAs have been implicated in the pathogenesis of cancer development being

involved in apoptosis, cell cycle control, cell proliferation and DNA repair. Studies have shown that the pattern of miRNA expression is altered in cancer and is different from that of the normal subjects⁷⁰⁻⁷² thus pointing towards their possible use as diagnostic markers. Studies indicate that a majority of miRNAs genes are located in cancer-associated genomic regions or in fragile sites.⁷³ The miRNAs have been shown to act as oncogenes (onco miRs) or tumour suppressors.⁷⁴ They have also been have a role in metastasis. These have been described as metastasis activators,⁷⁵ and their metastasis suppressor role has also been documented.⁷⁶

Dysregulation in miRNA expression have been reported in cancer of breast,⁷⁷ colon,⁷⁸ lung,⁷⁹ thyroid⁸⁰ etc. Circulating miRNAs are being recognized as potential biomarkers for the diagnosis of cancer.⁸¹⁻⁸³ Dysregulated expression of miR-125b, miR-145, miR-21, and miR-155 in breast cancer clearly differentiated normal from breast cancer tissues.⁷⁷ Down-regulation in miR-143 and miR-145 expression has been reported in colorectal cancer.⁷⁸ Up-regulation of 60 and down-regulation of 31 miRNAs in the serum from non-small cell lung cancer (NSCLC) patients compared to non-cancer (NC) patients has been reported.⁸⁴ Of these, a molecular signature of 4 miRNAs namely miR-193b, miR-301, miR-141 and miR-200b could discriminate lung cancer patients with high accuracy from NC patients. Recently the expression profile of seven miRNAs in serum of patients with papillary thyroid cancer (PTC), multinodular goiter (MG) and healthy controls has been reported.⁸⁵ The authors reported significantly higher levels of miR-21 in the pre-operative samples of PTC and MG patients compared to the control group. The MiR-151-5p, miR-221 and miR-222 were found to be significantly lower in the postoperative samples obtained from PTC patients. Their findings

imply the role of specific miRNAs in the development of PTC.⁸⁵

Future of circulating miRNAs

Thus, with increasing knowledge of the role of miRNAs in disease process and the discovery of circulating miRNAs in various diseases, a new branch of diagnostics based on molecular signatures of different miRNAs in diseases is coming up. The challenges' in this field would be to pinpoint disease-specific miRNAs and focus on methods to standardize collection techniques, analysis and interpretation of molecular signatures. Research in identifying potential miRNAs as therapeutic targets will be another new area in the over expanding field of microRNAs in coming years.

REFERENCES

1. Ladomery MR, Maddocks DG, Wilson ID. MicroRNAs: their discovery, biogenesis, function and potential use as biomarkers in non-invasive prenatal diagnostics. *Int J Mol Epidemiol Genet* 2011;2:253-60.
2. Mao XF, YC Cao. MicroRNA analysis in model species based on evolutionary rates. *Genet Mol Res* 2016;15:gm.15017216.
3. Cortez MA, Welsh JW, Calin G A. Circulating microRNAs as noninvasive biomarkers in breast cancer. *Recent Results Cancer Res* 2012;195:151-61.
4. Gomes P C, Cho H , Hood L, Franco OL, Pereira RW, Wang K. A review of computational tools in microRNAs discovery. *Front Genet* 2013;4:81.
5. Chalfie M, Horvitz HR, Sulston JE. Mutations that lead to reiterations in the cell lineages of *C. elegans*. *Cell* 1981;24:59-69.
6. Horvitz HR, Sulston JE. Isolation and genetic characterization of cell-lineage mutants of the nematode *Caenorhabditis elegans*. *Genetics* 1980;96:435-54.
7. Ferguson EL, Sternberg PW, Horvitz HR. A genetic pathway for the specification of the vulval cell lineages of *Caenorhabditis elegans*. *Nature* 1987;326:259-67.
8. Lee R, Feinbaum R, Ambros V. A short history of a short RNA. *Cell* 2004;116:S89-92.

9. Reinhart BJ, Slack FJ, Basson M, Pasquinelli AE, Bettinger JC, Rougvie AE, et al. The 21-nucleotide let-7 RNA regulates developmental timing in *Caenorhabditis elegans*. *Nature* 2000;403:901-6.
10. Lagos-Quintana M, Rauhut R, Lendeckel W, Tuschl T. Identification of novel genes coding for small expressed RNAs. *Science* 2001;294:853-8.
11. Pasquinelli AE, Reinhart BJ, Slack F, Martindale MQ, Kuroda MI, Maller B, et al. Conservation of the sequence and temporal expression of let-7 heterochronic regulatory RNA. *Nature* 2000;408:86-9.
12. Vrijens K, Bollati V and Navvrot T S. MicroRNAs as potential signatures of environmental exposure or effect: a systematic review. *Environ Health Perspect* 2015;123:399-411.
13. Allegra A, Alonci A, Campo S, Penna G, Petrungaro A, Gerace D, et al. Circulating microRNAs: New biomarkers in diagnosis, prognosis and treatment of cancer (review). *Int J Oncol* 2012;41:1897-912.
14. Zhou H, Huang X, Cui H, Luo X, Tang Y, Chen S, et al. miR-155 and its star form partner miR-155m cooperatively regulate type 1 interferon production by human plasma cytotid dendritic cells. *Blood* 2010;116:5885-94.
15. Hu HY, Yan Z, Xu Y, Hu H, Menzel C, Zhou YH, et al. Sequence features associated with microRNA strand selection in humans and flies. *BMC Genomics* 2009;10:413.
16. Fabian MR, Sonenberg N, Filipowicz W. Regulation of mRNA translation and stability by microRNAs. *Annu Rev Biochem* 2010;79:351-79.
17. Beezhold KJ, Castranova V, Chen F. Microprocessor of miRNAs: regulation and potential for therapeutic intervention. *Mol Cancer* 2010;9:134.
18. Ro S, Oark C, Young D, Sanders KM, Yan W. Tissue-dependent paired expression of miRNAs. *Nucleic Acids Res* 2007;35:5944-53.
19. Bartel DP. MiRNAs: target recognition and regulatory functions. *Cell* 2009;136:215-233.
20. Gibbins DJ, Ciaudo C, Erhardt M, Voinnet O. Multivesicular bodies associate with components of miRNA effector complexes and modulate miRNA activity. *Nat Cell Biol* 2009;11:1143-9.
21. Valadi H, Ekstrom K, Bossios A, Sjostrand M, Lee JJ, Lotvall JO. Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat Cell Biol* 2007;9:654-9.
22. Iguchi H, Kosaka N, Ochiya T. Secretory microRNAs as a versatile communication tool. *Commun Integr Biol* 2010;3:478-81.
23. Camussi G, Deregibus MC, Bruno S, Cantaluppi V, Biancone L. Exosomes/microvesicles as a mechanism of cell-to-cell communication. *Kidney Int* 2010;78:838-48.
24. Xie Y, Todd NW, Liu Z, Zhan M, Fang H, Peng H, et al. Altered miRNA expression in sputum for diagnosis of non-small cell lung cancer. *Lung Cancer* 2010;67:170-6.
25. Tsujiura M, Ichikawa D, Komatsu S, Shiozaki A, Takeshita H, Kosuga T, et al. Circulating microRNAs in plasma of patients with gastric cancers. *Br J Cancer* 2010;102:1174-9.
26. Huang Z, Huang D, Ni S, Peng Z, Sheng W, Du X. Plasma microRNAs are promising novel biomarkers for early detection of colorectal cancer. *Int J Cancer* 2010;127:118-26.
27. Wu F, Guo NJ, Tian H, Marohn M, Gearhart S, Bayless TM, et al. Peripheral blood microRNAs distinguish active ulcerative colitis and Crohn's disease. *Inflamm Bowel Dis* 2011;17:241-50.
28. Schöler N, Langer C, Kuchenbauer F. Circulating microRNAs as biomarkers - true blood? *Genome Med* 2011;3:72.
29. Keller A, Leidinger P, Bauer A, Elsharawy A, Haas J, Backes C, et al. Toward the blood-borne miRNome of human diseases. *Nat Methods* 2011;8:841-33.
30. Thum T, Catalucci D, Bauersachs J. MicroRNAs: novel regulators in cardiac development and disease. *Cardiovasc Res* 2008 1;79:562-70.
31. Cordes KR, Sheehy NT, White MP, Berry EC, Morton SU, Muth AN, et al. miR-145 and miR-143 regulate smooth muscle cell fate and plasticity. *Nature* 2009; 460:705-10.
32. Boettger T, Beetz N, Kostin S, Schneider J, Krüger M, Hein L, et al. Acquisition of the contractile phenotype by murine arterial smooth muscle cells depends on the Mir143/145 gene cluster. *J Clin Invest* 2009;119:2634-47.

33. Zhao W, Zhao SP, Zhao YH. MicroRNA-143/-145 in cardiovascular diseases. *Bio Med Res Int* 2015;2015:531740.
34. Mishra PK, Tyagi N, Kumar M, Tyagi SC. MicroRNAs as a therapeutic target for cardiovascular diseases. *JCell Mol Med* 2009;13:778-89.
35. Fangfei Deng, Xinran Xu and Yi-Han Chen. The role of miR-1 in the heart: From cardiac morphogenesis to physiological function. *Human Genet Embryol* 2014;4:1.
36. Yang B, Lin H, Xiao J, Lu Y, Luo X, Li B, et al. The muscle-specific microRNA miR-1 regulates cardiac arrhythmogenic potential by targeting GJA1 and KCNJ2. *Nat Med* 2007;13: 486-91.
37. Fish JE, Santoro MM, Morton SU, Yu S, Yeh RF, Wythe JD, et al. miR-126 regulates angiogenic signaling and vascular integrity. *Dev Cel.* 2008;15:272-84.
38. Fernandes T, Baraúna VG, Negrão CE, Phillips MI, Oliveira EM. Aerobic exercise training promotes physiological cardiac remodeling involving a set of microRNAs. *Am J Physiol Heart Circ Physiol* 2015;309:H543-52.
39. Chen W, Qin C. General hallmarks of microRNAs in brain evolution and development. *RNA Biol* 2015;12:701-8.
40. Ardekani AM, Naeini MM. The role of microRNAs in human diseases. *Avicenna J Med Biotechnol* 2010;2:161-79.
41. Bian S, Xu TL, Sun T. Tuning the cell fate of neurons and glia by microRNAs. *Curr Opin Neurobiol* 2013;23:928-34.
42. Cheng LC, Pastrana E, Tavazoie M, Doetsch F. miR-124 regulates adult neurogenesis in the subventricular zone stem cell niche. *Nat Neurosci* 2009;12:399-408.
43. Luikart BW, Bensen AL, Washburn EK, Perederiy JV, Su KG, Li Y, et al. miR-132 mediates the integration of newborn neurons into the adult dentate gyrus. *PLoS One* 2011;6):e19077.
44. Pedersen ME, Snieckute G, Kagias K, Nehammer C, Multhaupt HA, Couchman JR, et al. An epidermal microRNA regulates neuronal migration through control of the cellular glycosylation state. *Science* 2013;341:1404-8.
45. Cao DD, Li L, Chan WY. MicroRNAs: Key regulators in the central nervous system and their implication in neurological diseases. *Int J Mol Sci* 2016;17: pii: E842.
46. Giusti SA, Vogl AM, Brockmann MM, Vercelli CA, Rein ML, Trümbach D, et al, MicroRNA-9 controls dendritic development by targeting REST. *Elife* 2014;3.
47. Jasińska M, Miśek J, Cymerman IA, Eński S, Kaczmarek L, Dziembowska M. miR-132 regulates dendritic spine structure by direct targeting of matrix metalloproteinase 9 mRNA. *Mol Neurobiol* 2016;53:4701-12.
48. Schratt GM, Tuebing F, Nigh EA, Kane CG, Sabatini ME, Kiebler M, et al. A brain-specific microRNA regulates dendritic spine development. *Nature* 2006; 439:283-9.
49. Nelson PT, Wang WX, Rajeev BW. MicroRNAs (miRNAs) in neurodegenerative diseases. *Brain Pathol* 2008;18:130-8.
50. Lukiw WJ. Micro RNA speciation in fetal, adult and Alzheimer's disease hippocampus. *Neuro report* 2007;18:297-300.
51. Lukiw WJ, Pogue AI. Induction of specific micro RNA (miRNA) species by ROS-generating metal sulfates in primary human brain cells. *J Inorg Biochem* 2007;101:1265-9.
52. Schaefer A, O'Carroll D, Tan CL, Hillman D, Sugimori M, Llinas R, et al. Cerebellar neurodegeneration in the absence of microRNAs. *J Exp Med* 2007;204:1553-8 .
53. Shi W, Du J, Qi Y, Liang G, Wang T, Li S, et al. Aberrant expression of serum miRNAs in schizophrenia. *J Psychiatr Res* 2012;46:198-204.
54. Cogswell JP, Ward J, Taylor IA, Waters M, Shi Y, Cannon B, et al. Identification of miRNA changes in Alzheimer's disease brain and CSF yields putative biomarkers and insights into disease pathways. *J Alzheimers Dis* 2008;14:27-41.
55. Sonkoly E, Wei T, Janson PC, Sääf A, Lundberg L, Tengvall-Linder M, et al. MicroRNAs: novel regulators involved in the pathogenesis of psoriasis? *PLoS One* 2007;2:e610.

56. Taganov KD, Boldin MP, Chang KJ, Baltimore D. NF-kappaB-dependent induction of microRNA miR-146, an inhibitor targeted to signaling proteins of innate immune responses. *Proc Natl Acad Sci U S A* 2006;103:12481-6.
57. Pivarcsi A, Meisgen F, Xu N, Stähle M, Sonkoly E. Changes in the level of serum microRNAs in patients with psoriasis after antitumour necrosis factor- α therapy. *Br J Dermatol* 2013;169:563-70.
58. Kanemaru H, Fukushima S, Yamashita J, Honda N, Oyama R, Kakimoto A, et al. The circulating microRNA-221 level in patients with malignant melanoma as a new tumor marker. *J Dermatol Sci.* 2011;61:187-93.
59. Inoue K, Jinnin M, Yamane K, Makino T, Kajihara I, Makino K, Honda N et al. Down-regulation of miR-223 contributes to the formation of Gottron's papules in dermatomyositis via the induction of PKC[. *Eur J Dermatol* 2013;23:160-7.
60. O'Connell RM, Chaudhuri AA, Rao DS, Baltimore D. *Proc Natl Acad Sci USA* 2009;106:7113-8.
61. Tili E, Michaille JJ, Cimino A, Costinean S, Dumitru CD, Adair B, et al. Modulation of miR-155 and miR-125b levels following lipopolysaccharide/TNF-alpha stimulation and their possible roles in regulating the response to endotoxin shock. *J Immunol* 2007;179:5082-9.
62. Baulina NM, Kulakova OG, Favorova OO. MicroRNAs: The role in autoimmune inflammation. *Acta Naturae* 2016;8:21-33.
63. O'Carroll D, Mecklenbrauker I, Das PP, Santana A, Koenig U, Enright AJ, et al. A Slicer-independent role for Argonaute 2 in hematopoiesis and the microRNA pathway. *Genes Dev* 2007;21:1999-2004.
64. Bronevetsky Y, Villarino AV, Easley CJ, Barbeau R, Barczak AJ, Heinz GA, et al. T cell activation induces proteasomal degradation of Argonaute and rapid remodeling of the microRNA repertoire. *J Exp Med* 2013;210:417-32.
65. Nakasa T, Miyaki S, Okubo A, Hashimoto M, Nishida K, Ochi M, et al. Expression of microRNA-146 in rheumatoid arthritis synovial tissue. *Arthritis Rheum* 2008;58:1284-92.
66. Pauley KM, Satoh M, Chan AL, Bubbs MR, Reeves WH, Chan EK. Upregulated miR-146a expression in peripheral blood mononuclear cells from rheumatoid arthritis patients. *Arthritis Res Ther* 2008;10:R101.
67. Alsaleh G, Suffert G, Semaan N, Juncker T, Frenzel L, Gottenberg JE, et al. Bruton's tyrosine kinase is involved in miR-346-related regulation of IL-18 release by lipopolysaccharide-activated rheumatoid fibroblast-like synoviocytes. *J Immunol* 2009;182:5088-97.
68. Dai Y, Huang YS, Tang M, Lv TY, Hu CX, Tan YH, et al. Microarray analysis of microRNA expression in peripheral blood cells of systemic lupus erythematosus patients. *Lupus* 2007;16:939-46.
69. Dai Y, Sui W, Lan H, Yan Q, Huang H, Huang Y. Comprehensive analysis of microRNA expression patterns in renal biopsies of lupus nephritis patients. *Rheumatol Int* 2008; 29:749-54.
70. Croce CM. Causes and consequences of microRNA dysregulation in cancer. *Nat Rev Genet* 2009;10:704-14.
71. Calin GA, Croce CM. MicroRNA signatures in human cancers. *Nat Rev Cancer* 2006;6: 857-66.
72. Chen X, Ba Y, Ma L, Cai X, Yin Y, Wang K et al. Characterization of microRNAs in serum: A novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res.* 2008, 18: 997-1006.
73. Reddy KB. MicroRNA (miRNA) in cancer. *Cancer Cell Int* 2015;15:38.
74. Nicoloso MS, Spizzo R, Shimizu M, Rossi S, Calin GA. MicroRNAs—the micro steering wheel of tumour metastases. *Nat Rev Cancer* 2009;9:293-302.
75. Ma L, Teruya-Feldstein J, Weinberg RA. Tumour invasion and metastasis initiated by microRNA-10b in breast cancer. *Nature* 2007;449:682-8.
76. Tavazoie SF, Alarcón C, Oskarsson T, Padua D, Wang Q, Bos PD, et al. Endogenous human microRNAs that suppress breast cancer metastasis. *Nature* 2008;451:147-52.
77. Iorio MV, Ferracin M, Liu CG, Veronese A, Spizzo R, Sabbioni S, et al. MicroRNA gene expression deregulation in human breast cancer. *Cancer Res.* 2005;65:7065-70.
78. Michael MZ, O' Connor SM, van Holst Pellekaan NG, Young GP, James RJ. Reduced accumulation of specific microRNAs in colorectal neoplasia. *Mol Cancer Res* 2003;1:882-91.

79. Takamizawa J, Konishi H, Yanagisawa K, Tomida S, Osada H, Endoh H, et al. Reduced expression of the let-7 microRNAs in human lung cancers in association with shortened postoperative survival. *Cancer Res* 2004;64:3753-6.
80. He H, Jazdzewski K, Li W, Liyanarachchi S, Nagy R, Volinia S, et al. The role of microRNA genes in papillary thyroid carcinoma. *Proc Natl Acad Sci U S A*. 2005;102:19075-80.
81. Mahn R, Heukamp LC, Rogenhofer S, von Ruecker A, Muller SC, Ellinger J. Circulating micro RNAs (miRNA) in serum of patients with prostate cancer. *Urology* 2011;77:1265.e9-16.
82. Bianchi F, Nicassio F, Marzi M, Belloni E, Dall'olio V, Bernard L, et al. A serum circulating miRNA diagnostic test to identify asymptomatic high-risk individuals with early stage lung cancer. *EMBO Mol Med* 2011;3:495-503.
83. Sun Y, Wang M, Lin G, Sun S, Li X, Qi J, et al. Serum microRNA-155 as a potential biomarker to track disease in breast cancer. *PLoS One* 2012;7:e47003.
84. Nadal E, Truini A, Nakata A, Lin J, Reddy RM, Chang AC, et al. A novel serum 4-microRNA signature for lung cancer detection. *Sci Rep* 2015;5:12464.
85. Yoruker EE, Terzioglu D, Teksoz S, Uslu FE, Gezer U, Dalay N. MicroRNA expression profiles in papillary thyroid carcinoma, benign thyroid nodules and healthy controls. *J Cancer* 2016;7:803-9.